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<u>L3</u>	L2 and antibod\$3	2	<u>L3</u>
<u>L2</u>	RI-H	12	<u>L2</u>
<u>L1</u>	RI-H near5 protein	0	<u>L1</u>

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=> s soluble lactose binding lectin

L1 40 SOLUBLE LACTOSE BINDING LECTIN

=> s l1 and human

L2 14 L1 AND HUMAN

=> s l1 and antibod###

L3 11 L1 AND ANTIBOD###

=> s l3 and (diagnos## or clinic## or in vivo or in vitro)

L4 0 L3 AND (DIAGNOS## OR CLINIC## OR IN VIVO OR IN VITRO)

=> s l3 and diagnos##

L5 0 L3 AND DIAGNOS##

=> dup rem l3

PROCESSING COMPLETED FOR L3

L6 4 DUP REM L3 (7 DUPLICATES REMOVED)

=> d l6 1-4 bib ab kwic

L6 ANSWER 1 OF 4 MEDLINE on STN DUPLICATE 1
AN 96185805 MEDLINE
DN 96185805 PubMed ID: 8604014
TI Regulated expression of a 16-kd galectin-like protein in activated rat macrophages.
AU Rabinovich G; Castagna L; Landa C; Riera C M; Sotomayor C
CS Departamentos de Bioquimica Clinica y Quimica Biologica, Facultad de Ciencias Quimicas, Universidad Nacional de Cordoba, Argentina.
SO JOURNAL OF LEUKOCYTE BIOLOGY, (1996 Mar) 59 (3) 363-70.
Journal code: 8405628. ISSN: 0741-5400.
CY United States
DT Journal; Article; (JOURNAL ARTICLE)
LA English
FS Priority Journals
EM 199605
ED Entered STN: 19960524
Last Updated on STN: 19960524
Entered Medline: 19960515
AB We investigated the presence of a galectin-like protein in rat mononuclear cells using a polyclonal **antibody** raised against a **soluble lactose-binding lectin** purified from adult chicken liver that immunoreacted strongly with a broad protein band of about 16 kd in Western blot assays. Immunochemical studies revealed a constitutive expression of this protein in mononuclear cells mainly in the macrophage (M phi) population. Subcellular localization was assessed by Western blot assays of the cytosolic and membrane fractions of different cell populations studied: (1) spleen mononuclear cells, (2) T cell-enriched, (3) B cell- and M phi-enriched populations, and (4) peritoneal cells, processed in the presence of lactose. In broad agreement with immunocytochemical studies of

nonpermeabilized and permeabilized cells, Western blot assays suggest that this protein is localized mainly in the cytoplasmic compartment but also associated with the cell surface. By flow cytometric analyses we detected about a 14% of ED1 double-positive cells corresponding to macrophages that constitutively express this galectin-like protein associated with their cell surface. The cytosolic fraction obtained from the M phi-enriched cell population showed hemagglutinating activity specifically inhibited by beta-galactoside-related sugars. Moreover, this galectin-like protein was retained in a lactosyl-Sepharose matrix and specifically eluted with lactose. In this work, evidence is also provided to show that different stimuli are able to modulate the expression of the galectin-like protein. Expression was upregulated in inflammatory and activated macrophages, revealing a significant increase in phorbol ester- and formylmethionine oligopeptide-treated cells. Both stimuli involving protein kinase C activation pathway have been able not only to up-regulate the total expression of this protein but also to modulate its subcellular localization.

AB We investigated the presence of a galectin-like protein in rat mononuclear cells using a polyclonal **antibody** raised against a **soluble lactose-binding lectin** purified from adult chicken liver that immunoreacted strongly with a broad protein band of about 16 kd in Western blot. . .

L6 ANSWER 2 OF 4 MEDLINE on STN DUPLICATE 2
 AN 95095551 MEDLINE
 DN 95095551 PubMed ID: 8002251
 TI Distribution of an endogenous 16-kd S-lac lectin in the chicken retina.
 AU Castagna L F; Landa C A
 CS Department of Biological Chemistry, Faculty of Chemical Sciences, Universidad Nacional de Cordoba, Argentina.
 SO INVESTIGATIVE OPHTHALMOLOGY AND VISUAL SCIENCE, (1994 Dec) 35 (13) 4310-6. Journal code: 7703701. ISSN: 0146-0404.
 CY United States
 DT Journal; Article; (JOURNAL ARTICLE)
 LA English
 FS Priority Journals
 EM 199501
 ED Entered STN: 19950215
 Last Updated on STN: 19950215
 Entered Medline: 19950125

AB PURPOSE. To examine by indirect immunofluorescence the distribution of an endogenous 16-kd S-lac lectin (**soluble lactose binding lectin**) during development of the chicken retina. METHODS. Cryosections of retinal tissue at different developmental stages and cultured retinal cells (either not permeabilized or permeabilized with acetone) were incubated with a rabbit antiserum that specifically reacts with the retinal 16-kd S-lac lectin. After incubation with a fluorescent-labeled secondary **antibody**, tissue sections and cultured cells were analyzed by fluorescence microscopy. RESULTS. Retina was weakly stained with the antiserum on early embryonic day 7, whereas on embryonic days 13 and 18 it showed a restricted "granular" staining in the outer retina. At embryonic day 18, in addition, there was widespread staining in all retinal layers. This pattern was maintained by postnatal day 5 and in the adult retina, although the intensity of the staining of the outer retina was weaker. In retinal cell cultures, glial-like flat cells and monopolar, bipolar, and multipolar neurons were stained with the antiserum, but only if they had been previously permeabilized with acetone. CONCLUSION. The results suggest that the distribution of a 16-kd S-lac lectin changes during retinal development. Cell culture experiments indicate that most often the lectin is localized intracellularly in the different retinal cell types.

AB PURPOSE. To examine by indirect immunofluorescence the distribution of an endogenous 16-kd S-lac lectin (**soluble lactose binding lectin**) during development of the chicken

retina. METHODS. Cryosections of retinal tissue at different developmental stages and cultured retinal cells (either. . . incubated with a rabbit antiserum that specifically reacts with the retinal 16-kd S-lac lectin. After incubation with a fluorescent-labeled secondary **antibody**, tissue sections and cultured cells were analyzed by fluorescence microscopy. RESULTS. Retina was weakly stained with the antiserum on early. . .

CT Check Tags: Animal; Support, Non-U.S. Gov't
Cells, Cultured
Chick Embryo
Chickens
Cross Reactions
Electrophoresis, Polyacrylamide Gel
Fluorescent Antibody Technique
Immunoblotting
*Lactose: ME, metabolism
*Lectins: ME, metabolism
Molecular Weight
Retina: EM, embryology
Retina: GD, growth & development

L6 ANSWER 3 OF 4 MEDLINE on STN DUPLICATE 3
AN 92268105 MEDLINE
DN 92268105 PubMed ID: 1375225
TI Isolation and expression of a gene encoding L-14-II, a new human **soluble lactose-binding lectin**.
AU Gitt M A; Massa S M; Leffler H; Barondes S H
CS Department of Psychiatry, Langley Porter Psychiatric Institute, University of California, San Francisco 94143.
SO JOURNAL OF BIOLOGICAL CHEMISTRY, (1992 May 25) 267 (15) 10601-6.
Journal code: 2985121R. ISSN: 0021-9258.
CY United States
DT Journal; Article; (JOURNAL ARTICLE)
LA English
FS Priority Journals
OS GENBANK-L01141; GENBANK-M87010; GENBANK-M87842; GENBANK-M87857; GENBANK-M87858; GENBANK-M87859; GENBANK-M87860; GENBANK-S96735; GENBANK-S96741; GENBANK-S96751; GENBANK-S96754
EM 199206
ED Entered STN: 19920710
Last Updated on STN: 19970203
Entered Medline: 19920625
AB In the course of screening a human hepatoma cDNA library with **antibody** raised against a mammalian lectin with subunit molecular weight of about 14,000, we detected a partial cDNA encoding a related but distinct protein that was possibly a homologous lectin (Gitt and Barondes, 1986). We here report the isolation and sequencing of a full-length cDNA for this protein from a HepG2 cDNA library. The cDNA encodes a protein with subunit molecular weight of 14,650. Expression of the coding sequence in Escherichia coli yields a product that binds to a lactose affinity column and is specifically eluted with lactose, confirming that this new protein is a lectin. Like its well studied relative, here called L-14-I, the new lectin, L-14-II, exists as a homodimer in solution. The two related human lectins have 43% amino acid sequence identity. The genomic DNA encoding L-14-II (LGALS2) contains four exons with similar intron placement to L-14-I (LGALS1); but the genomic upstream region, which contains several sequences characteristic of regulatory elements, differs significantly from L-14-I.
TI Isolation and expression of a gene encoding L-14-II, a new human **soluble lactose-binding lectin**.
AB In the course of screening a human hepatoma cDNA library with **antibody** raised against a mammalian lectin with subunit molecular weight of about 14,000, we detected a partial cDNA encoding a related. .

L6 ANSWER 4 OF 4 MEDLINE on STN
 AN 90243778 MEDLINE
 DN 90243778 PubMed ID: 2335567
 TI Evidence for export of a muscle lectin from cytosol to extracellular matrix and for a novel secretory mechanism.
 AU Cooper D N; Barondes S H
 CS Department of Psychiatry, University of California, San Francisco 94143-0984.
 NC 1-RO1-HL38627 (NHLBI)
 SO JOURNAL OF CELL BIOLOGY, (1990 May) 110 (5) 1681-91.
 Journal code: 0375356. ISSN: 0021-9525.
 CY United States
 DT Journal; Article; (JOURNAL ARTICLE)
 LA English
 FS Priority Journals
 OS GENBANK-X51903
 EM 199006
 ED Entered STN: 19900706
 Last Updated on STN: 19900706
 Entered Medline: 19900608
 AB **A soluble lactose-binding lectin**
 with subunit Mr of 14,500 is believed to function by interacting with extracellular glycoconjugates, because it has been detected extracellularly by immunohistochemistry. This localization has been questioned, however, since the lectin lacks a secretion signal sequence, which challenges the contention that it is secreted. We have demonstrated externalization of this lectin from C2 mouse muscle cells by both immunoprecipitation of metabolically labeled protein and immunohistochemical localization. We further show that externalization of the lectin is a developmentally regulated process that accompanies myoblast differentiation and that the lectin codistributes with laminin in myotube extracellular matrix. Immunohistochemical localization during intermediate stages of externalization suggests that the lectin becomes concentrated in evaginations of plasma membrane, which pinch off to form labile lectin-rich extracellular vesicles. This suggests a possible mechanism for lectin export from the cytosol to the extracellular matrix.
 AB **A soluble lactose-binding lectin**
 with subunit Mr of 14,500 is believed to function by interacting with extracellular glycoconjugates, because it has been detected extracellularly. . . .
 CT Check Tags: Animal; Support, Non-U.S. Gov't; Support, U.S. Gov't, Non-P.H.S.; Support, U.S. Gov't, P.H.S.
 Amino Acid Sequence
 Antibody Specificity
 Base Sequence
 Biological Transport: PH, physiology
 Cell Differentiation: PH, physiology
 *Cytosol: ME, metabolism
 Exocytosis: PH, physiology
 *Extracellular Matrix:. . .

=> s RI-H (10a) protein#
 L7 8 RI-H (10A) PROTEIN#

=> s l7 and antibod###
 L8 4 L7 AND ANTIBOD###

=> s l8 and human
 L9 0 L8 AND HUMAN

=> s l8 and cancer

L10 0 L8 AND CANCER

=> dup rem l8

PROCESSING COMPLETED FOR L8

L11 1 DUP REM L8 (3 DUPLICATES REMOVED)

=> d l11 bib ab kwic

L11 ANSWER 1 OF 1 MEDLINE on STN DUPLICATE 1
AN 95347007 MEDLINE
DN 95347007 PubMed ID: 7621528
TI Rat intestinal galactoside-binding lectin L-36 functions as a structural protein in the superficial squamous cells of the esophageal epithelium.
AU Wasano K; Hirakawa Y
CS Department of Anatomy, Faculty of Medicine, Kyushu University, Fukuoka, Japan.
SO CELL AND TISSUE RESEARCH, (1995 Jul) 281 (1) 77-83.
Journal code: 0417625. ISSN: 0302-766X.
CY GERMANY: Germany, Federal Republic of
DT Journal; Article; (JOURNAL ARTICLE)
LA English
FS Priority Journals
EM 199508
ED Entered STN: 19950911
Last Updated on STN: 19950911
Entered Medline: 19950825
AB Using an affinity purified **antibody** raised against the **RI-H** fragment of rat intestinal lectin L-36, the latter **protein** has been identified within the esophageal epithelium by means of ultracryotomy followed by immunogold labeling. The epithelium consists of 4 morphologically distinct cell-types, namely, the basal, spiny, granular and squamous cells, and each of these exhibits a different immunolabeling pattern. The basal cells form a layer on the basal lamina, and in these a diffuse cytoplasmic staining is observed. This basal cell layer is overlaid by spiny cells that extend many cell processes into wide intercellular spaces. In these cells, immunogold particles are found only on small granular inclusions consisting of an electron-lucent homogeneous substance. The granular cells form a third layer over the spiny cells, and are characterized by a number of large granular inclusions with an electron-dense core rimmed by a less electron-dense substance. Immunogold labeling is found on these granules, both on the core and peripheral region. Squamous cell-types constitute the most superficial layer of the epithelium. They are without granular inclusions, and immunogold labeling is confined to the cytoplasmic surface of the thickened plasma membrane. These findings suggest that L-36 is produced in the basal cells as free cytosolic protein, then becomes progressively aggregated into the granular inclusions of the spiny and granular cells, and is eventually transferred onto the cytoplasmic surface of the squamous cell plasma membrane where it may interact with complementary glycoconjugate(s) located at this site. (ABSTRACT TRUNCATED AT 250 WORDS)
AB Using an affinity purified **antibody** raised against the **RI-H** fragment of rat intestinal lectin L-36, the latter **protein** has been identified within the esophageal epithelium by means of ultracryotomy followed by immunogold labeling. The epithelium consists of 4. . .

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